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## Recent developments and advances in atopic dermatitis and food allergy

Sugita, Kazunari ; Akdis, Cezmi A

**Abstract:** This review highlights recent advances in atopic dermatitis (AD) and food allergy (FA), particularly on molecular mechanisms and disease endotypes, recent developments in global strategies for the management of patients, pipeline for future treatments, primary and secondary prevention and psychosocial aspects. During the recent years, there has been major advances in personalized/precision medicine linked to better understanding of disease pathophysiology and precision treatment options of AD. A greater understanding of the molecular and cellular mechanisms of AD through substantial progress in epidemiology, genetics, skin immunology and psychological aspects resulted in advancements in the precision management of AD. However, the implementation of precision medicine in the management of AD still requires the validation of reliable biomarkers, which will provide more tailored management, starting from prevention strategies towards targeted therapies for more severe diseases. Cutaneous exposure to food via defective barriers is an important route of sensitization to food allergens. Studies on the role of the skin barrier genes demonstrated their association with the development of IgE-mediated FA, and suggest novel prevention and treatment strategies for type 2 diseases in general because of their link to barrier defects not only in AD and FA, but also in asthma, chronic rhinosinusitis, allergic rhinitis and inflammatory bowel disease. The development of more accurate diagnostic tools, biomarkers for early prediction, and innovative solutions require a better understanding of molecular mechanisms and the pathophysiology of FA. Based on these developments, this review provides an overview of novel developments and advances in AD and FA, which are reported particularly during the last two years.

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## Invited Review Article

## Recent developments and advances in atopic dermatitis and food allergy

Kazunari Sugita <sup>a, b, c</sup>, Cezmi A. Akdis <sup>a, b, \*</sup><sup>a</sup> Swiss Institute of Allergy and Asthma Research (SIAF), University of Zurich, Davos Platz, Switzerland<sup>b</sup> Christine Kühne-Center for Allergy Research and Education, Davos, Switzerland<sup>c</sup> Division of Dermatology, Department of Medicine of Sensory and Motor Organs, Tottori University Faculty of Medicine, Yonago, Japan

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## ABSTRACT

This review highlights recent advances in atopic dermatitis (AD) and food allergy (FA), particularly on molecular mechanisms and disease endotypes, recent developments in global strategies for the management of patients, pipeline for future treatments, primary and secondary prevention and psychosocial aspects. During the recent years, there has been major advances in personalized/precision medicine linked to better understanding of disease pathophysiology and precision treatment options of AD. A greater understanding of the molecular and cellular mechanisms of AD through substantial progress in epidemiology, genetics, skin immunology and psychological aspects resulted in advancements in the precision management of AD. However, the implementation of precision medicine in the management of AD still requires the validation of reliable biomarkers, which will provide more tailored management, starting from prevention strategies towards targeted therapies for more severe diseases. Cutaneous exposure to food via defective barriers is an important route of sensitization to food allergens. Studies on the role of the skin barrier genes demonstrated their association with the development of IgE-mediated FA, and suggest novel prevention and treatment strategies for type 2 diseases in general because of their link to barrier defects not only in AD and FA, but also in asthma, chronic rhinosinusitis, allergic rhinitis and inflammatory bowel disease. The development of more accurate diagnostic tools, biomarkers for early prediction, and innovative solutions require a better understanding of molecular mechanisms and the pathophysiology of FA. Based on these developments, this review provides an overview of novel developments and advances in AD and FA, which are reported particularly during the last two years.

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## Introduction

There are remarkable recent developments understanding the mechanisms, diagnostic and therapeutic management of atopic dermatitis (AD) and food allergy (FA) and differences in the practice by dermatologists and pediatricians in different countries. AD is a chronic inflammatory pruritic skin disease that affects a large number of children and adults in industrialized countries. Its worldwide prevalence, ranges from 0.2% to 24.6%, with the highest prevalence of AD in childhood in Africa and Latin America.<sup>1</sup> The onset of AD occurs during the first year of life in 60% of the children.<sup>2</sup> The prevalence of AD in the first 2 years of life is 21.5%.<sup>3</sup> 43.2% of these children have spontaneous outgrow of the disease, 38.3%

persist with mild AD and mild rhinitis and 18.7% persist with severe disease and show an extensive atopic march including FA, asthma, and rhinitis.<sup>3</sup> Only 16.8% of AD patients have onset after adolescence.<sup>4</sup> AD and FA are extensively overlapping. The prevalence of FA has been suggested to increase in the last two decades and is in the range of 4% in food challenge tests in 1 year, which increases to 6–8% in questionnaire surveys.<sup>5</sup> FA shows various different clinical presentations with respect to responsible allergens, age at presentation, timing of reaction, presence of comorbid atopic diseases, outgrow and resolution with time, and response to immunotherapy. This heterogeneity of clinical presentations of FA possess a challenge to successful management and treatment. Avoidance of allergenic foods and the use of epinephrine in case of a severe reaction triggered by accidental ingestion remain the standard of care, as there are currently no approved treatments for FA. Dysregulated immune response patterns and their heterogenous and complex combination in chronic inflammation, immune cell and tissue cell hyperresponsiveness, microinflammation even at the

\* Corresponding author. University of Zurich, Swiss Institute of Allergy and Asthma Research, Davos Platz, Switzerland.

E-mail address: [akdisac@siaf.uzh.ch](mailto:akdisac@siaf.uzh.ch) (C.A. Akdis).

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healing stage and tissue remodeling in affected tissues define the complexity of FA and AD.

Modern healthcare is rapidly developing and staying away from the concept of including all of the patients in one basket to an individualized response with the combination of precision diagnosis and personalized treatment. An overarching medicinal concept is developing with better understanding of genotypes, phenotypes, endotypes, regiotypes, biomarkers and theratypes of diseases that also fully include AD and FA.<sup>6</sup> More than 100 years old allergen-specific management of allergic diseases has particularly contributed to the early awareness in precision medicine. Multi-pleomics, big data, and systems biology have demonstrated a profound complexity and dynamic variability in AD and FA between individuals, as well as between regions.<sup>6</sup>

Basic research has intensified during the recent years to better understand the immune and inflammatory mechanisms in AD and FA. IgE sensitization, essential molecular pathways of type 2 response, mechanisms of disease outgrow and therapy response are important in both diseases. Mechanisms of desensitization, successful ways of treatment without side effects and short- and long-term tolerance mechanisms are key research areas in FA. This article highlights and summarizes key advances in AD and FA published during the last couple of years. Important ones from these achievements are summarized in [Tables 1 and 2](#).

### Global strategies for the assessment and management of atopic dermatitis

There is a consistent association of AD with other atopic diseases including asthma and FA. The concept of atopic march was developed to describe the progression of these atopic diseases in children.<sup>7</sup> Although it most often starts in infancy, it is also highly prevalent in adults.<sup>8</sup> Among the adult AD population, a multidimensional burden has been described not only including dermatologic symptoms, but also increased socioeconomic burden, sleep disorders, and reductions

in health-related quality of life and work productivity.<sup>9–11</sup> With increasing severity, the multidimensional burdens listed above also increase. International Classification of Diseases, 11th Revision (ICD-11) was provided,<sup>12</sup> and ICD-9, clinical modification codes alone were found to be insufficient for identification of AD from healthcare databases, but incorporation of the diagnosis of asthma, hay fever, and FA improved the positive predictive value and specificity of these searches.<sup>13</sup> The Harmonizing Outcomes Measures for Eczema (HOME) initiative is an evidence driven and evidence-generating outcomes research initiative that aims to develop standardization of a core set of outcome measurements of atopic eczema, which is also known as AD or eczema.<sup>14,15</sup> Various instruments exist to measure symptoms in AD to assess safety and efficacy of therapies in clinical trials.<sup>16,17</sup> Among 18 instruments, the pediatric Itch Severity Scale (ISS), Patient-Oriented Eczema Measure (POEM), Patient-Oriented SCORing Atopic Dermatitis (PO-SCORAD), Self-Administered Eczema Area and Severity Index (SA-EASI), and adapted SA-EASI are currently recommended for assessing symptoms of AD in future clinical trials.<sup>18</sup> However, the use of simple patient-reported global AD severity scores is feasible for clinical practice and epidemiological research. A simple scoring system of mild, moderate, and severe patient-reported AD severity correlated well with objective AD assessments, objective SCORAD, and EASI.<sup>19</sup> Patient-reported AD severity may be sufficiently valid for assessing AD severity. For the management of AD, patient's increased adherence to topical medication is indispensable particularly with use of topical corticosteroids. The topical corticosteroid phobia (TOPICOP) score is available for the assessment of topical corticosteroid phobia based on 12 items.<sup>20</sup> TOPICOP scorings can feasibly be applied internationally and can be used to obtain data from studies and to adapt patient education and treatment.<sup>21</sup>

The lifetime prevalence of atopic diseases is significantly increasing from adolescence to adulthood, particularly that of allergic rhinitis.<sup>22</sup> Regarding the clinical course of AD, no significant difference in AD prevalence before and after childhood has been found by systemic review and meta-analysis of longitudinal

**Table 1**  
Key developments and advances in atopic dermatitis (AD).

Developments and Advances	References
<b>Global strategies for the assessment and management of AD</b>	
Validation of International Classification of Disease	Hsu <i>et al.</i> <sup>13</sup>
Evaluation of symptom measurement instruments	Blauvelt <i>et al.</i> <sup>17</sup> , Vakharia <i>et al.</i> <sup>19</sup> , Stalder <i>et al.</i> <sup>21</sup>
Lifetime prevalence	Mortz <i>et al.</i> <sup>22</sup> , Abuabara <i>et al.</i> <sup>23</sup> , Barbarot <i>et al.</i> <sup>24</sup>
Product-specific standardization	Zimmer <i>et al.</i> <sup>26</sup> , Bonertz <i>et al.</i> <sup>28</sup>
<b>Prevention of AD</b>	
Atopic march	Han <i>et al.</i> <sup>29</sup> , Ferreira <i>et al.</i> <sup>31</sup>
Microbiome and immune system	Chu <i>et al.</i> <sup>35</sup>
Prevention approach to decrease AD	Elbert <i>et al.</i> <sup>38</sup> , Roduit <i>et al.</i> <sup>41</sup> , Silverwood <i>et al.</i> <sup>43</sup>
<b>Psychological aspects of AD</b>	
Management of psychological status	Ronnstad <i>et al.</i> <sup>48</sup>
Attention-deficit/hyperactivity disorder	Schmitt <i>et al.</i> <sup>53</sup>
<b>Mechanisms and pathophysiology of AD</b>	
microRNA and AD	Carreras-Badosa <i>et al.</i> <sup>60</sup>
Periostin and epidermal barrier	Mitamura <i>et al.</i> <sup>61</sup> , Izuhara <i>et al.</i> <sup>62</sup>
Biomarker	Czarnowicki <i>et al.</i> <sup>65</sup>
T-cell receptor repertoire	Brunner <i>et al.</i> <sup>66</sup>
Mast cells and sphingosine-1-phosphate activation	Wedman <i>et al.</i> <sup>69</sup>
Langerhans and inflammatory dendritic epidermal cells	Otsuka <i>et al.</i> <sup>72</sup>
Aryl hydrocarbon receptor mediated anti-inflammatory feed back	Koch <i>et al.</i> <sup>75</sup>
<b>Therapeutic targets and experimental models for the treatment of AD</b>	
Allergen specific immunotherapy	Shin <i>et al.</i> <sup>131</sup>
Oral tolerance	Baek <i>et al.</i> <sup>77</sup>
Topical ivermectin	Ventre <i>et al.</i> <sup>79</sup>
Mesenchymal stem cells	Song <i>et al.</i> <sup>80</sup>
Antimicrobial peptide	Kopfngel <i>et al.</i> <sup>83</sup>
Chemokine receptors	Murray <i>et al.</i> <sup>85</sup>
BCG vaccination	Thostesen <i>et al.</i> <sup>87</sup>

**Table 2**

Key developments and advances in food allergy (FA).

Developments and Advances	References
<b>Diagnosis and the role of biomarkers in early prediction</b>	
Precision medicine in FA	Agache <i>et al.</i> <sup>6</sup> , Flores <i>et al.</i> <sup>97</sup>
Assessing basophil activation	Mukai <i>et al.</i> <sup>95</sup>
Prediction of FA	Datema <i>et al.</i> <sup>100</sup> , Pettersson <i>et al.</i> <sup>109</sup> , Baumert <i>et al.</i> <sup>111</sup>
Predictive biomarkers of anaphylaxis	Turner <i>et al.</i> <sup>112</sup>
<b>Innovative solutions</b>	
Allergen exposure chambers	Rosner-Friese <i>et al.</i> <sup>124</sup> , Pfaar <i>et al.</i> <sup>126</sup>
Companion animal research	Jensen-Jarolim <i>et al.</i> <sup>134</sup>
<b>Mechanisms and pathophysiology</b>	
Vitamin D and mast cell stabilization	Liu <i>et al.</i> <sup>145</sup>
Epigenetic mechanisms	Asai <i>et al.</i> <sup>150</sup>
Intestinal microbiome and food sensitization	Savage <i>et al.</i> <sup>149</sup>
microRNA and FA	D'Argenio <i>et al.</i> <sup>153</sup>
Tolerance development	Sampson <i>et al.</i> <sup>147</sup>
Characterization of the T-cell response	Zulehner <i>et al.</i> <sup>164</sup>
<b>Novel strategies for the treatment and management</b>	
Allergen immunotherapy	Burks <i>et al.</i> <sup>166</sup> , Wood <sup>171</sup> , Ryan <i>et al.</i> <sup>174</sup>
Patient's quality of life	Rigbi <i>et al.</i> <sup>168</sup>
Nanoparticles for allergen-specific immunotherapy	Pohlit <i>et al.</i> <sup>176</sup>
Combination therapy	MacGinnitie <i>et al.</i> <sup>173</sup>
Early peanut and egg introduction	Greenhawt <i>et al.</i> <sup>180</sup> , Shaker <i>et al.</i> <sup>182</sup>

studies.<sup>23</sup> In studies of AD prevalence by country, estimations of adult AD prevalence ranges from 2.1% to 4.9%.<sup>24</sup> For international clinical and epidemiological research, it is becoming increasingly clear that differing nomenclature of the disease has important harmful or unwanted consequences.<sup>25</sup> AD as well as the term atopic eczema describe clinically chronic relapsing pruritic inflammatory skin conditions. In this context, the International Eczema Council (IEC) noted that the term eczema is imprecise and confusing and recommended the use of AD or atopic eczema in all publications, presentations and discussions about the disorder.<sup>25</sup>

Product-specific standardization is of critical importance for patients with suspected or proven allergy.<sup>26</sup> Ensuring persistent quality, safety, and efficacy of medical allergen products is required for an effective treatment. Products for allergen immunotherapy (AIT) have been approved by national authorities; however, different regulations exist worldwide. Although regulatory framework in the European Union (EU) has been developed in the field of AIT, it remains heterogeneous. New allergen products for AIT are being developed and manufactured each year. However, despite the increased number of medical products such as AIT, regulatory framework for quality parameters has not yet been reached. Thus, it is extremely complicated and challenging to develop a harmonized international approach to regulation. International pharmaceutical companies have recently placed an emphasis on medical products that promote health and supply local and neighboring markets as well as global markets. Therefore, it is quite important to understand the regulation of allergen products from a global perspective. From these kind of circumstances, the EAACI Taskforce on Regulatory Aspects of Allergen Immunotherapy, as a part of the EAACI AIT Guidelines, analyzed how products for the diagnosis of allergies and AIT are regulated in different countries and regions worldwide.<sup>27</sup> They also provided an overview of how AIT products are regulated with respect to their manufacturing and quality in EU, and United States.<sup>28</sup> They described similarities and differences in the regulation of allergen manufacturing and quality control between the EU and United States.

### Prevention of atopic dermatitis

The development of AD in infancy and subsequent conditions such as FA, asthma, and allergic rhinitis has been observed as co-manifestation or occurring in sequence. These longitudinal

associations have been known to lead to atopic march,<sup>29</sup> and an association between AD and birch pollen allergy has also been reported.<sup>30</sup> There has been controversy regarding whether atopic march is the primary causal factor in childhood allergic diseases. A recent study indicated that AD, asthma and allergic rhinitis partly coexist, because they share many genetic risk variants that dysregulate the expression of immune-related genes.<sup>31</sup> Another study showed that the risk of developing asthma or allergic rhinitis was much higher in IgE-associated AD compared to nonallergic AD.<sup>32</sup> Therefore, it has been argued that it is important to distinguish between eczema with and without IgE sensitization when considering the implications of allergic diseases in infancy.<sup>32,33</sup>

Environmental factors, such as bacteria influence the human microbiome and immune system. Development of novel preventative approaches is important, and a recent review summarizes current evidence for the potential of bacteria and their metabolites in the prevention of allergic diseases.<sup>34</sup> Early life microbial exposure influences the risk of allergies and development of a balanced immune system.<sup>35,36</sup> Not only the exposure, but also the correct composition of the gut microbiota have a role in allergy development.<sup>37</sup>

Furthermore, the increased incidence of AD and the progression from AD to allergic rhinitis and asthma have highlighted the need for disease prevention. A recent article indicates that shorter duration or non-exclusive breastfeeding was associated with a weak overall increased risk of AD, but not sensitization.<sup>38</sup> Concerning the influence of pregnancy-related and perinatal factors, children who are born by cesarean section and assisted-delivery showed significant associations with AD, asthma and atopic sensitization in childhood.<sup>39</sup> Effect of food diversity in childhood has also been investigated. The consumption of cheese had a significant protective effect for AD.<sup>40</sup> The effect on AD may be associated with the diversity of consumed cheese, including those rich in microbial diversity. It has to be noted here that cheese is an important provider of short chain fatty acids that are linked to prevention of AD in children and mouse models.<sup>41</sup> Despite a preventative approach to decrease AD in early-childhood, some patients may have persistent disease into adulthood. However, the prevalence of comorbidities in patients with AD is not well characterized.<sup>42</sup> Recent studies have shown that patients with severe AD had a high prevalence of smoking, stroke, cardiovascular disease, inflammatory bowel disease, depression and anxiety.<sup>42,43</sup>

These comorbidities might represent targets for prevention and intervention of AD management.

### Psychological aspects of atopic dermatitis

AD has been thought to carry an increased risk for mental disorders.<sup>8</sup> However, few studies have reported the prevalence of psychiatric disorders among adults with AD. A recent study revealed that adolescents with AD from Korea are associated with a higher prevalence of depression symptoms and suicidal behaviors.<sup>44</sup> A poor health-related quality of life is linked with depressive mood, depression, and suicidal ideation, which are, in turn, associated with AD.<sup>45</sup> Therefore, improving poor health-related quality of life and managing psychological status in AD among adolescents are critical. From a population-based survey using large-scale observational data, association among depression, anxiety, and AD has been analyzed.<sup>46</sup> The risk of hospitalization and suicide was also examined. They found a significant association between self-reported AD among adults and clinician-diagnosed psychiatric status, including depression and anxiety.<sup>46</sup> Moreover, a recent study of the association between AD and suicidality suggests that patients with AD are at an increased risk of suicidal ideation and suicide attempts.<sup>47</sup> However, depression, anxiety, and suicidal ideation do not lead to psychiatric consultations, hospitalization, or suicide.<sup>46</sup> One study reported that improvement of AD appears to be reduced these psychiatric conditions.<sup>48</sup>

Epidemiological studies using standardized diagnostic criteria suggest that 3%–6% of the child population may suffer from attention-deficit/hyperactivity disorder (ADHD) and more recently, a relevant association between AD and ADHD has been reported.<sup>49,50</sup> Moreover, children with ADHD are more likely to have not only AD but asthma, allergic rhinitis, and allergic conjunctivitis than their counterparts.<sup>51</sup> A Swedish birth cohort study found that preschool eczema was not associated with ADHD medication at school age.<sup>52</sup> To shed more light on the comorbidity of AD and ADHD, Schmitt *et al.* hypothesized that children with AD, who do not meet the complete diagnostic criteria of AD frequently show features of ADHD and increased mental health problems.<sup>53</sup> They concluded that even if the clinical diagnosis of ADHD is excluded, children with AD show increased levels of ADHD symptoms.<sup>53</sup> Despite these findings, the underlying biological mechanisms still require further investigation. Therefore, prevention and effective treatment of AD represent unmet needs in children as well.

### Mechanisms and pathophysiology of atopic dermatitis

Major mechanisms of AD include abnormalities in the terminal differentiation of keratinocytes that lead to a defective stratum corneum.<sup>54,55</sup> A defective barrier in AD allows the penetration of allergens and microbes, leading to IgE sensitization and type 2 cytokine production that drives allergic inflammation.<sup>56</sup> At least two AD subtypes can be distinguished: the first type is characterized by a strong type 2 mediated response with high levels of serum immunoglobulin E (IgE) and the second is a nonallergic type characterized with normal IgE levels.<sup>8,57</sup> MicroRNAs (miRNAs) are short, single-stranded RNA molecules that regulate Th2 skewing and influence innate and adaptive immune responses.<sup>58,59</sup> Among miRNAs, miRNA-146a is needed for the production of IgE and linked to the modulation of Th1/Th17 cell mediated responses in mice.<sup>60</sup> A matricellular protein, periostin, is induced by Th2 cytokines IL-4 or IL-13 and plays an important role in barrier dysfunction. A recent study shows that IL-24, produced in keratinocytes downstream of periostin and IL-13, decrease filaggrin expression that leads to barrier dysfunction in AD.<sup>61,62</sup> In AD, Th2 cells as well as other T-cell subpopulations (Th1, Th17, and Th22) are detectable

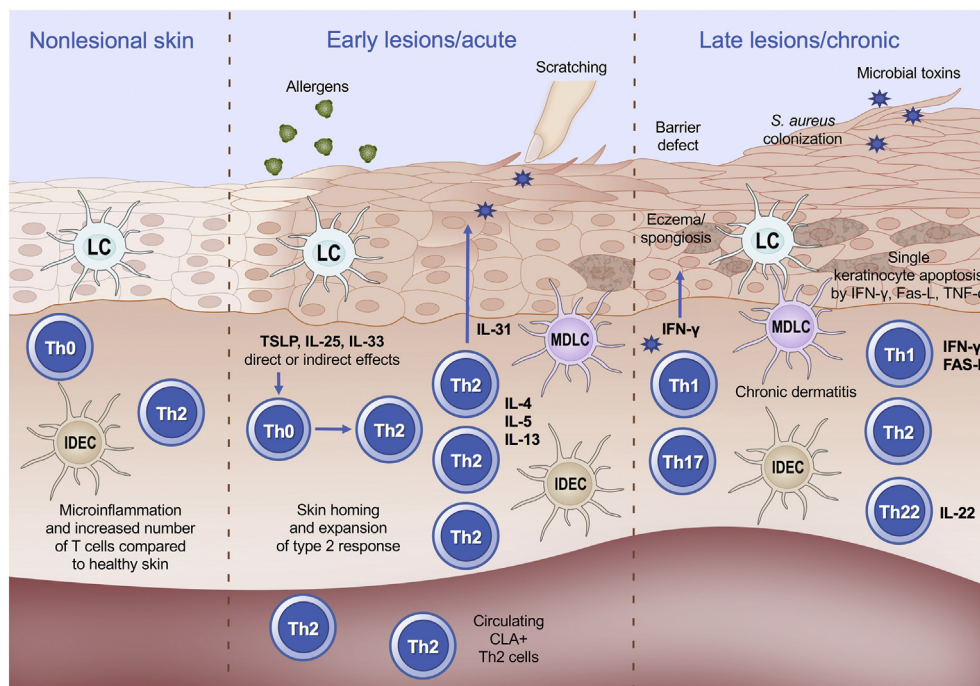
in the AD skin lesions. Figure 1 shows immune and inflammatory cells in AD. It is important to emphasize that the majority of T cells in AD skin lesions are cutaneous lymphocyte-associated antigen (CLA<sup>+</sup>)CD45RO<sup>+</sup> memory T cells.<sup>63,64</sup> Due to recirculation, peripheral CLA<sup>+</sup>T cells have similar features characteristic of T cells present in AD skin lesions.<sup>65</sup> Various skin T cells, such as CD4 TH1 cells, CD4 TH2, CD4 TH17, CD4 TH22, CD4 Treg, resident memory T cells,  $\gamma\delta$  T cells, CD8 TC1, CD8 TC2, Mucosal-associated invariant T cells (MAIT) are described in Table 3. One recent advancement in AD was evidence showing that circulating CLA<sup>+</sup> T cells can be a reliable peripheral biomarker of inflammatory events occurring in the skin.<sup>65</sup> There is conflicting literature regarding the diversity of T-cell receptor (TCR) repertoire in AD skin lesions and their relationship to nonlesional skin. In this context, Brunner *et al.* showed that nonlesional AD skin shares a TCR repertoire similar to lesional AD skin.<sup>66</sup>

Other cell types, such as innate lymphoid cells (ILC), mast cells, eosinophils, basophils, and inflammatory dendritic cell populations can be detected in AD skin.<sup>67</sup> In human skin mast cells, proteomic analysis identifies two novel mast cell proteins, L1CAM/CD171 and DPP4/CD26, that are potentially relevant to skin homeostasis.<sup>68</sup> Skin mast cell and sphingosine-1-phosphate (S1P) activation is a novel effector that initiates remodeling in AD.<sup>69</sup> Several ILC-modulating factors are dysregulated in AD and the levels of IL-25, IL-33, TSLP, and PGD2 are elevated in AD skin.<sup>70</sup> In addition, cell–cell interactions between ILC2s and keratinocytes lead to activation or suppression of ILC2s.<sup>70</sup> Identification of surface molecules on these cells involved in type 2 immunity could provide new therapeutic targets for the treatment of AD. Blom *et al.* identified, for the first time, the expression of CD200R by cells such as ILC2, Th2, and basophils. Interleukin-31 (IL-31), preferentially produced from Th2 cells, binds to the IL-31 receptor (IL-31R) expressed on sensory nerves. IL-31/IL31R signaling has recently been shown to play a critical role in the development of pruritus in AD.<sup>71</sup> In the skin, there exists at least three populations of antigen-presenting cells including Langerhans cells (LCs), monocyte-derived LC-like cells, and inflammatory dendritic epidermal cells (IDECs).<sup>72</sup> IDECs and LC-like cells have been found to be present in both steady states and inflammatory states, and they are present in AD.<sup>73</sup> LC and IDEC in AD skin do not respond to Toll-like receptor (TLR) 2 activation and may contribute to the inability to clear *S. aureus* infection.<sup>74</sup> LC in patients with AD carry the high-affinity receptor for IgE, Fc $\epsilon$ RI, and are engaged in the pathogenesis of AD. The aryl hydrocarbon receptor mediates the anti-inflammatory feedback mechanism in Fc $\epsilon$ RI-expressing human LC.<sup>75</sup>

### Future therapeutic targets and experimental models for the treatment of atopic dermatitis

Although there are many available treatments for allergic diseases, AIT induces establishment of long-term clinical tolerance to allergens, resulting in the prevention of further development of allergic diseases and a role for primary prevention has also been suggested.<sup>76</sup> Among currently investigated AIT routes in FA, oral immunotherapy (OIT) involves the oral administration of increasing amounts of allergens.<sup>76</sup> The transcriptomic profiles of skin obtained from mice that were epicutaneously sensitized, but orally tolerized indicate that oral antigen administration provides protection against AD by the expression of genes regulating Th2 inflammatory responses and skin barrier function.<sup>77,78</sup> In a relevant mouse model of AD, another study reported the efficacy of topical ivermectin, a drug used for scabies and rosacea, improved AD by inhibiting the priming and activation of allergen-specific T cells.<sup>79</sup> Several recent studies have shown promising results in the use of mesenchymal stem cells for the treatment of AD.<sup>80,81</sup> Sah *et al.*





**Fig. 1.** T cells and dendritic cells in atopic dermatitis. T cells in uninvolved skin, early lesions and chronic lesions of type 2 AD. T cells are moderately increased in the dermis in the dermis of non-lesional skin. Skin homing CLA<sup>+</sup> T cells display a Th2 profile in early lesions and in the circulation. However, in the course of the disease, increasing populations of Th1 cells and IFN- $\gamma$  and sometimes Th17 and Th22 cells are detected in chronic lesions of AD. IL-33, IL-25 and TSLP released from keratinocytes directly or indirectly support the type 2 response. IL31 released from T cells plays a major role in itch. Three populations of antigen-presenting cells including Langerhans cells (LCs) monocyte-derived LC-like cells, and inflammatory dendritic epidermal cells (IDECs) exist in the skin. IDECs and LC-like cells have been found to be present in both steady states and inflammatory states, and they are present in lesional AD.

**Table 3**

T cell subsets in the skin.

Different conventional and non-conventional T cells	Major functions
CD4 TH1	AD chronic lesions, ACD, drug exanthemas, mechanisms of eczema formation
CD4 TH2	AD lesions, AD and general allergy allergen-specific Th2, eosinophilic skin disease
CD4 TH17	AD chronic lesions, psoriasis, neutrophilic drug exanthemas
CD4 TH22	AD late lesions
CD4 Treg	Controls overactivation of other subsets in peripheral blood and lymphatic organs in AD, ACD. Does not exist too many in lesional skin
T <sub>RM</sub>	Resident memory T cells, mainly against infectious agents, mostly CD8
$\gamma\delta$ T cells	Against infectious agents in mice, very few in human skin
CD8 TC1	Against viruses and other infectious agents, found in AD and ACD skin and drug exanthemas
CD8 TC2	AD, eosinophilic skin diseases
MAIT cells	Mucosal-associated invariant T cells, against bacteria and fungi, riboflavin responsive and MR-1 tetramer responsive

AD, atopic dermatitis; ACD, allergic contact dermatitis.

explored the clinical potential for superoxide dismutase 3-transduced mesenchymal stem cells in mice with AD. They showed that subcutaneously administrated superoxide dismutase 3-transduced mesenchymal stem cells suppress skin inflammation via the histamine H4 receptor/IL-4 receptor-dependent mechanism.<sup>81</sup> Succinate is an intermediate of the citric acid cycle that binds to its specific receptor, SUCNR1/GPR91. In addition, it acts as an alarmin and is involved in tissue injury or inflammatory stimulus. Furthermore, GPR91, a metabolic control receptor for the binding of succinate, deficiency in mice leads to allergic contact dermatitis, suggesting that GPR91 is a therapeutic target for the treatment of allergic skin diseases.<sup>82</sup>

In humans, Kopfnagel *et al.* investigated the effect of RNase 7, a 14.5-kDa antimicrobial ribonuclease on isolated human T cells.<sup>83</sup> They demonstrated that CD4<sup>+</sup> T cells from AD patients showed a less pronounced downregulation of IL-13 in response to RNase 7 compared to healthy controls.<sup>83</sup> Thus, RNase 7 has immunoregulatory functions on Th2 cells and reduces the production of Th2 cytokines in the skin.

Many receptors have been studied, such as CCR4, CCR10, and CCR8, for the recruitment of immune cells to the skin.<sup>84–86</sup> Due to the complexity of AD, it would be interesting to target specific cells with cocktails of chemokine receptor antagonists administered as ointments on the skin.<sup>84</sup> In a randomized clinical trial of neonatal

BCG vaccination in the Danish population, BCG vaccination at birth was found to be associated with less AD among children with atopic predisposition, although more studies are needed to confirm these findings.<sup>87</sup>

### Precision medicine for food allergy: diagnosis and the role of biomarkers in early prediction

Molecular mechanism-linked therapies can be defined as therapies tailored to the characteristics of each patient to obtain a better clinical outcome. These different therapeutic responses linked to individual patient's biological mechanisms is known as precision medicine.<sup>88</sup> Precision medicine consists of four main components including disease taxonomy, digital monitoring of patients and biomarkers, disease phenotypes and endotypes, and biomarker- and endotype-linked patient care and therapies.<sup>89</sup> Three main pillars of precision medicine are endotypes, phenotypes, and biomarkers.<sup>89</sup> Important concepts such as regiotypes and theratypes have been recently introduced. Both regiotypes (regional differences in pheno and endotypes as well as allergens affecting different parts of the world) and theratypes (treatment response) represent important advancements for precision diagnosis and treatment.<sup>6</sup> Of all allergic diseases, FA is particularly well-suited as a target for precision medicine. This is due to its pathophysiology, the modulation of which has resulted in better clinical outcomes, and additionally has been clearly associated with detectable biomarkers.<sup>90,91</sup> The consensus document of EAACI and American Academy of Allergy, Asthma and Immunology (AAAAI) has been recently published under the auspices of the PRACTALL collaboration platform.<sup>92</sup> PRACTALL is an initiative of EAACI and AAAAI to harmonize the European and American approach to allergic diseases with regard to patient treatment and scientific progress.<sup>90</sup> In the consensus document, special consideration was given to defining endotypes in order to assist in developing novel therapeutic approaches. The definition of endotypes in FA is necessary to develop novel therapeutic approaches. FA endotypes can be characterized into IgE-mediated endotypes, including alpha-gal allergy and oral allergy syndrome, and non-IgE-related and mixed endotypes such as food protein-induced gastrointestinal endotype and eosinophilic endotype.<sup>93,94</sup> Diagnostic tools and biomarkers for accurate diagnosis, evaluation of prognosis, and efficacy of treatment are still under investigation, but basophil activation test and component-resolved diagnostics are promising tools.<sup>95,96</sup> A recent systematic review showed that selected components of cow's milk, hen's egg, peanut, hazelnut, and shrimp allergen revealed high specificity for diagnosis of FA in component resolved diagnosis.<sup>97,98</sup> In a multicenter study, the detection of Ana o 3-specific IgE provides a predictive value in the diagnosis in children with suspected cashew allergy.<sup>99</sup> A combination of component-resolved diagnostics and other factors such as clinical background and extract-based serology has been proposed to predict severe reactions to hazelnut allergy.<sup>100</sup> Several biomarkers have been associated with FA including skin prick tests, allergen specific IgE, IgG4, genetic, and epigenetic markers.<sup>92,101–104</sup> A study of patients with meat allergy revealed that the IgG subclass distribution in these patients is distinct from natural IgG responses in nonallergic individuals.<sup>105</sup> Due to a frequent co-sensitization to birch pollen allergens and profilins, diagnosis of lipid transfer proteins (LTPs) using plant extracts is difficult.<sup>106</sup> In a study from a central European population, Pru p 3, peach LTP, can be used as an allergen marker for LTP sensitization.<sup>107</sup> To identify diagnostic markers for anaphylaxis in food allergic patients, Wittenberg *et al.* found serum levels of apolipoproteins as a useful biomarker of anaphylaxis.<sup>108</sup>

Early prediction of the severity of FA is one of the key issues for the accurate diagnosis and improved management. The extent to

which the severity of food allergic reactions can be predicted by clinical factors such as age, type of food, and eliciting dose is currently unknown.<sup>109</sup> Several studies provided evidence that the eliciting dose contributes to the frequency of accidental reactions.<sup>110,111</sup> Another study suggests that individuals have a threshold of reactivity for symptoms in general as well as a threshold for symptoms specifically of anaphylaxis.<sup>112</sup> Additionally, a double-blind, placebo controlled study on patients with food challenge-confirmed FA to milk, egg, peanut, cashew, and/or hazelnut indicated that clinicians should not use the eliciting dose obtained from a graded food challenge.<sup>109</sup> This is because eliciting dose only contributes to reaction severity. A successful diagnostic allergy work-up is required to tailor medical treatments to individual characteristics of each FA patient.<sup>113</sup> Extensive immune and allergy workups are becoming more and more important for many diseases.<sup>114</sup> For FA diagnosis, a comprehensive toolbox for improved documentation and decision-making using oral food challenges has been provided.<sup>115</sup> With these tools, we hope to reduce the influence of subjective judgment of supervising physicians.<sup>115</sup>

### Innovative solutions: allergen exposure chambers

Allergen exposure chambers have proven to be valuable tools for diagnosis and determining safety and efficacy of new therapeutics.<sup>116–119</sup> This tool can help to develop further evidence for allergen immunotherapy as well as biological therapies, which can potentially control allergic diseases.<sup>120–122</sup> Allergen exposure chambers also provide a new setting which has generated evidence that in patients with AD with grass pollen sensitization is linked to a worsening of cutaneous symptoms.<sup>123</sup> Although facilities have been reported as technically validated, various technical setups and specifications are found in allergen exposure chamber facilities.<sup>124</sup> Furthermore, several unmet needs require attention and evaluation before allergen exposure chambers can be used in allergen immunotherapy clinical trials.<sup>125</sup> Thus, it is necessary to standardize allergen exposure chambers, and harmonize protocols for clinical and immunological research in order to improve data quality and facilitate better application of allergen exposure chambers. Currently, the EAACI position paper harmonizes current concepts of allergen exposure chambers with regard to standardization of the challenge procedure and assessments, pediatric issues, and regulatory issues.<sup>126</sup> These relevant unmet needs and issues may be addressed before determining safety and efficacy of new therapeutics.<sup>126</sup>

### Animal models in AD and FA

Animal models of human diseases are commonly utilized for the development of new drugs and the analysis of pathogenic research. Current animal research targets specific immune mechanisms involved in human atopic and allergic diseases.<sup>127</sup> In AD, AIT has been recently reported in a randomized controlled trial and meta-analysis, especially in patients sensitized to the house dust mite antigen.<sup>128–130</sup> However, there is still controversy about the role of AIT as a therapeutic intervention in AD. Several reasons exist for this, such as there is no relevant mouse model to investigate the mechanism and validate AIT in AD and primary end points were not achieved in several proof of concept clinical trials in AD. Shin *et al.* established an AIT model of AD using *Dermatophagoides farinae* treated NC/Nga mice and demonstrated clinical and histological improvement.<sup>131</sup> They found that induction of Tregs and IL-10-producing NK cells is a possible source of IL-10 and this model may be a useful tool to analyze the efficacy of AIT modalities for the treatment of AD.<sup>131</sup>

In a recent study a method to directly assess *in vivo* the epidermal barrier function by electrical impedance (EI) spectroscopy was first time demonstrated in mice demonstrating a reliable diagnostic tool to detect skin barrier defects.<sup>132</sup> Mice epidermal barrier was damaged by epicutaneous application of proteases and cholera toxin and by tape stripping. EI and transepidermal water loss (TEWL) were measured before and after the application. A few hours after papain application, a dose-dependent reduction of EI was detected, reflecting the decreased barrier function. 24 and 48 h after the treatment, EI starts to increase to background levels, indicating tissue healing and restoration of skin barrier.<sup>132</sup> Barrier disruption was confirmed by histological analysis showing an impaired stratum corneum and higher cellular infiltration after papain application.<sup>132</sup> In addition, immunofluorescence staining and RT-PCR showed downregulation of molecules involved in the barrier function, such as filaggrin, occludin and claudin-1 and mRNA levels of filaggrin, loricrin, and involucrin. Comparable results were observed after tape stripping and cholera toxin treatment.

In FA, various animals have been used for studying the mechanisms of FA.<sup>133</sup> Adverse reactions to food also occur in companion animals and comparison between humans and animal models may lead to improved recommendations for the prevention and treatment of FA.<sup>134</sup> Similar to humans, companion animals may spontaneously develop skin, respiratory, and gastrointestinal tract symptoms.<sup>135</sup> However, there is no clear evidence between adverse food reactions and respiratory symptoms in companion animals.<sup>136</sup> For example, in dogs, adverse food reactions may manifest as cutaneous symptoms, which frequently are indistinguishable from AD due to environmental allergens.<sup>137</sup> In addition, little is known about the mechanisms of FA reactions in companion animals. In dogs, an allergen specific IgE response and CD8<sup>+</sup> T cell phenotype have been studied for the mechanism of FA.<sup>138</sup> Based on the current knowledge and the EAACI position paper, comparative studies of companion animals and humans suffering from FA may serve to fill knowledge gaps in this area.<sup>136</sup>

### Mechanisms and pathophysiology of food allergy

In food allergic patients, exposure to food antigens elicits a type 1 hypersensitivity reaction, which induces anaphylaxis. Atopic diseases are significantly associated with food induced anaphylaxis both in children and adults, but not with anaphylaxis induced by drug and venom.<sup>139</sup> Oral tolerance is the mechanism by which we maintain a normal physiological response to food antigens, and the breakdown of oral tolerance is likely to be linked to sensitization to food allergens. In humans, early skin barrier disruption due to inflammation is associated with an increased risk of FA.<sup>140,141</sup> Environmental exposure to food allergens can occur in house and bed dust after hen's egg consumption.<sup>142</sup> Importantly, oral exposure is also one of the routes for sensitization to allergens.<sup>143</sup> Recently, Lexmond *et al.* demonstrated that a loss of function mutation in the Wiskott-Aldrich syndrome (Was) protein lead to an increased risk of FA.<sup>144</sup> They showed that Was<sup>-/-</sup> mice became sensitized to OVA upon oral administration in the absence of adjuvant, resulting in the development of IgE-mediated mast cell responses and FA. Studies from Was<sup>-/-</sup> mice will provide new insights into the pathophysiology of FA in humans. Vitamin D plays a critical role in the stabilization of mast cells, which automatically activate in vitamin D deficient conditions.<sup>145</sup> However, there is no evidence that vitamin D insufficiency during the first 6 month of infancy is a risk factor for IgE-mediated FA at 1 year of age.<sup>146</sup> Other factors such as genes (e.g., filaggrin gene mutation) and microbiome are also related to FA.<sup>147</sup> One potentially important gene is the signal transducer and activator of transcription 6 (STAT6),

which stimulates Th2 differentiation. In this context, van Ginkel *et al.* identified for the first time that risk variants increase STAT6 expression and STAT6 variants may be involved in the pathogenesis of FA.<sup>148</sup> Multiple studies have provided evidence that FA is determined in part, by the microbiome.<sup>141</sup> Prior studies investigated the association in infants of the intestinal microbiome and allergic disease in humans.<sup>37</sup> In a prospective microbiome-wide association study, bacterial genera present in the infant intestinal microbiome were inversely associated with the development of food sensitization at the age of 3 years.<sup>149</sup> Recently, epigenetic mechanisms have been suggested to play a role in FA.<sup>147,150</sup> For example, differences in epigenetic changes in DNA methylation of the promoter region of Th1 and Th2 cytokine genes and Treg play a role in children with IgE-mediated cow's milk allergy.<sup>151,152</sup> Identification of specific miRNAs and their targets may help to understand the mechanism of FA. miR193a-5p is the most downregulated miRNA in children with cow's milk allergy and the inhibition of miR193a-5 resulted in elevation of protein level of IL-4.<sup>153</sup>

Although allergy to cow's milk and hen egg is more common among younger children, the mechanisms of the natural development of tolerance remains poorly understood.<sup>154</sup> They are typically sensitized with cow's milk proteins, such as casein and lactoglobulin.<sup>155</sup> IgG4 antibodies have been proposed to be a good indicator of the clinical efficacy of allergy treatments.<sup>156</sup> Recent analysis of IgG4 epitope binding in patients with cow's milk allergy in children revealed that the number of peptides bound by IgG4 antibodies and intensity of binding decreased when patients became tolerant, suggesting that these antibodies are important in tolerance acquisition.<sup>157</sup> Cow's milk formulas differ with regard to the degree of hydrolysis of the milk protein. In this context, several mild hydrolyzed cow's formulas are available and useful for the avoidance of cow's milk allergy in children due to different properties with regard to allergenic activity, IgE reactivity, T-cell proliferation, and cytokine secretion.<sup>158</sup> Tolerance to baked egg and cow's milk is related to conformational changes induced by heating.<sup>159</sup> However, reaction threshold data from 352 children, who underwent food challenges to fresh or extensively heated hen's egg and cow's milk found no significant shift in dose-distribution curves by baking process.<sup>160</sup>

Almost 70% of patients with birch pollen sensitivity have IgE-mediated reactions to plant foods such as hazelnut, apple, stone fruits, kiwi, and carrot, which typically cause oral allergy syndrome.<sup>161,162</sup> These food allergies are caused by a sensitization to Bet v 1 and subsequent IgE and T-cell cross-reactivity to food allergens.<sup>161</sup> Dau c 1 has been suggested as a major allergy in carrot allergic patients.<sup>163</sup> Dau c 1-specific T cell lines established from patients with birch pollen and carrot allergy indicated that T cell response to Dau c1 occurs independently of Bet v 1, suggesting that in addition to cross-reactivity with the major pollen allergens, Dau c 1 itself shows sensitization activity.<sup>164</sup>

### Novel strategies for the treatment and management of food allergy

The current treatment approach of FA is strict avoidance of the offending food or foods and the use of rescue medication in the event of an allergic reaction.<sup>165</sup> Therefore, a significant amount of research has been conducted on food AIT including OIT, sublingual immunotherapy (SLIT), and epicutaneous immunotherapy (EPIT). The aim of food AIT is to achieve clinical desensitization, sustained unresponsiveness, and oral tolerance, which are essential for emerging therapies for FA.<sup>166</sup> The EAACI Task Force on Allergen Immunotherapy for IgE-mediated FA provided evidence-based recommendations for active treatment with AIT.<sup>167</sup> Food AIT has shown the greatest promise for children 4–5 years of age with



symptoms suggestive of persistent IgE-mediated FA to cow's milk, hen's egg, or peanut.<sup>167</sup> In addition to the improvement of symptoms, quality of life of patients with FA improves during OIT for allergy.<sup>168</sup> In other words, a greater understanding of psychological and physical outcomes contribute to prevention and intervention in FA.<sup>169</sup> Current studies have enrolled patients with heterogeneous clinical symptoms.<sup>170</sup> Most children with milk and egg allergy develop tolerance spontaneously. Although OIT has been shown to be more efficient than SLIT, OIT has demonstrated a higher frequency of adverse events.<sup>167</sup> In clinical settings, increasing doses during the build-up phase should be performed in food AIT.<sup>171</sup> Novel studies have been conducted adding anti-IgE as a biological to improve food AIT. Addition of anti-IgE therapy reduces the allergic side effects of food AIT as it was shown for other AITs.<sup>172,173</sup> To ensure efficacy and safety of food AIT, patient factors such as age, co-existence of other allergic diseases, and adherence should be considered.<sup>167,170</sup> Development of education and training of primary care professionals, diagnosis and stratification of patients, and monitoring of treatment effectiveness are required for AIT.<sup>174</sup>

New developments in technology and the combination of two therapeutic strategies offers opportunities to improve treatment with AIT.<sup>175</sup> Encapsulation of allergens or DNA vaccines into nanoparticles may provide advantages compared to conventional AIT.<sup>176,177</sup> Srivastava *et al.*, investigated the efficacy and safety of peanut OIT using CpG-coated nanoparticles containing a peanut extract in a mouse model of peanut allergy.<sup>178</sup> They found that mice treated with the nanoparticles were significantly protected from anaphylaxis.<sup>178</sup> For the improvement of SLIT treatment, combination therapy with systemic administration of IL-2/anti-IL-2 complexes suppresses experimental FA in mice. Combination therapy may represent a promising strategy for the treatment and management of FA.<sup>179</sup> Another recent focus has turned to primary prevention of FA.<sup>180</sup> Early peanut introduction is one way to prevention, but early peanut recommendations differ among countries with formal guidelines.<sup>181</sup> Furthermore, there is evidence supporting that early egg introduction can decrease the risk of developing egg allergy.<sup>182</sup>

## Conclusion

In conclusion, we have reviewed the recent developments on AD and FA. An important feature of these recent developments is that the era of precision medicine brings novel strategies in the management and treatment of AD and FA. Development of a prevention strategy, early interventions and psychological disturbance are important aspects of patient care. The emergence of novel diagnostic tools, innovative solutions, and biomarkers as well as novel mechanisms of AD and FA can be used to accelerate the development of novel drugs and targeted therapies. Identifying treatment effects and looking at candidate biomarkers in both animal models and humans will provide evidence to promote the ability to practice personalized treatment.

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## Conflict of interest

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## References

1. Odhiambo JA, Williams HC, Clayton TO, Robertson CF, Asher MI, Group IPTS. Global variations in prevalence of eczema symptoms in children from ISAAC Phase Three. *J Allergy Clin Immunol* 2009;**124**:1251–8.
2. Kay J, Gawkrödger DJ, Mortimer MJ, Jaron AG. The prevalence of childhood atopic eczema in a general population. *J Am Acad Dermatol* 1994;**30**:35–9.
3. Illi S, von Mutius E, Lau S, Nickel R, Gruber C, Niggemann B, et al. The natural course of atopic dermatitis from birth to age 7 years and the association with asthma. *J Allergy Clin Immunol* 2004;**113**:925–31.
4. Ozkaya E. Adult-onset atopic dermatitis. *J Am Acad Dermatol* 2005;**52**:579–82.
5. Dunlop JH, Keet CA. Epidemiology of food allergy. *Immunol Allergy Clin North Am* 2018;**38**:13–25.
6. Agache I, Akdis CA. Precision medicine and phenotypes, endotypes, genotypes, regiotypes, and theratypes of allergic diseases. *J Clin Invest* 2019;**130**:1493–503.
7. Bantz SK, Zhu Z, Zheng T. The atopic march: progression from atopic dermatitis to allergic rhinitis and asthma. *J Clin Cell Immunol* 2014;**5**.
8. Weidinger S, Novak N. Atopic dermatitis. *Lancet* 2016;**387**:1109–22.
9. Chang YS, Chiang BL. Sleep disorders and atopic dermatitis: a 2-way street? *J Allergy Clin Immunol* 2018;**142**:1033–40.
10. Eckert L, Gupta S, Amand C, Gadkari A, Mahajan P, Gelfand JM. Impact of atopic dermatitis on health-related quality of life and productivity in adults in the United States: an analysis using the National Health and Wellness Survey. *J Am Acad Dermatol* 2017;**77**:274–279. e3.
11. Chung J, Simpson EL. The socioeconomics of atopic dermatitis. *Ann Allergy Asthma Immunol* 2019;**122**:360–6.
12. Tanno LK, Calderon M, Linzer JF, Chalmers RJ, Demoly P; Joint Allergy Academies. Collaboration between specialties for respiratory allergies in the international classification of diseases (ICD)-11. *Respir Res* 2017;**18**:34.
13. Hsu DY, Dalal P, Sable KA, Voruganti N, Nardone B, West DP, et al. Validation of international classification of disease ninth revision codes for atopic dermatitis. *Allergy* 2017;**72**:1091–5.
14. Schmitt J, Apfelbacher C, Spuls PI, Thomas KS, Simpson EL, Furue M, et al. The Harmonizing Outcome Measures for Eczema (HOME) roadmap: a methodological framework to develop core sets of outcome measurements in dermatology. *J Invest Dermatol* 2015;**135**:24–30.
15. Schmitt J, Spuls P, Boers M, Thomas K, Chalmers J, Rookevisch E, et al. Towards global consensus on outcome measures for atopic eczema research: results of the HOME II meeting. *Allergy* 2012;**67**:1111–7.
16. Beck LA, Thaci D, Hamilton JD, Graham NM, Bieber T, Rocklin R, et al. Dupilumab treatment in adults with moderate-to-severe atopic dermatitis. *N Engl J Med* 2014;**371**:130–9.
17. Blauvelt A, de Bruin-Weller M, Gooderham M, Cather JC, Weisman J, Pariser D, et al. Long-term management of moderate-to-severe atopic dermatitis with dupilumab and concomitant topical corticosteroids (LIBERTY AD CHRONOS): a 1-year, randomised, double-blinded, placebo-controlled, phase 3 trial. *Lancet* 2017;**389**:2287–303.
18. Gerbens LA, Prinsen CA, Chalmers JR, Drucker AM, von Kobyletzki LB, Limpens J, et al. Evaluation of the measurement properties of symptom measurement instruments for atopic eczema: a systematic review. *Allergy* 2017;**72**:146–63.
19. Vakharia PP, Chopra R, Sacotte R, Patel N, Immaneni S, White T, et al. Validation of patient-reported global severity of atopic dermatitis in adults. *Allergy* 2018;**73**:451–8.
20. Aubert-Wastiaux H, Moret L, Le Rhun A, Fontenoy AM, Nguyen JM, Leux C, et al. Topical corticosteroid phobia in atopic dermatitis: a study of its nature, origins and frequency. *Br J Dermatol* 2011;**165**:808–14.
21. Stalder JF, Aubert H, Anthoine E, Futamura M, Marcoux D, Morren MA, et al. Topical corticosteroid phobia in atopic dermatitis: international feasibility study of the TOPICOP score. *Allergy* 2017;**72**:1713–9.
22. Mortz CG, Andersen KE, Poulsen LK, Kjaer HF, Broesby-Olsen S, Bindsvlev-Jensen C. Atopic diseases and type I sensitization from adolescence to adulthood in an unselected population (TOACS) with focus on predictors for allergic rhinitis. *Allergy* 2019;**74**:308–17.
23. Abuabara K, Yu AM, Okhovat JP, Allen IE, Langan SM. The prevalence of atopic dermatitis beyond childhood: a systematic review and meta-analysis of longitudinal studies. *Allergy* 2018;**73**:696–704.
24. Barbarot S, Auziere S, Gadkari A, Girolomoni G, Puig L, Simpson EL, et al. Epidemiology of atopic dermatitis in adults: results from an international survey. *Allergy* 2018;**73**:1284–93.
25. Silverberg JI, Thyssen JP, Paller AS, Drucker AM, Wollenberg A, Lee KH, et al. What's in a name? Atopic dermatitis or atopic eczema, but not eczema alone. *Allergy* 2017;**72**:2026–30.
26. Zimmer J, Vieths S, Kaul S. Standardization and regulation of allergen products in the European Union. *Curr Allergy Asthma Rep* 2016;**16**:21.
27. Bonertz A, Roberts GC, Hoefnagel M, Timon M, Slater JE, Rabin RL, et al. Challenges in the implementation of EAACI guidelines on allergen immunotherapy: a global perspective on the regulation of allergen products. *Allergy* 2018;**73**:64–76.
28. Bonertz A, Roberts G, Slater JE, Bridgewater J, Rabin RL, Hoefnagel M, et al. Allergen manufacturing and quality aspects for allergen immunotherapy in Europe and the United States: an analysis from the EAACI AIT Guidelines Project. *Allergy* 2018;**73**:816–26.

29. Han H, Roan F, Ziegler SE. The atopic march: current insights into skin barrier dysfunction and epithelial cell-derived cytokines. *Immunol Rev* 2017;**278**:116–30.
30. Wassmann-Otto A, Heratizadeh A, Wichmann K, Werfel T. Birch pollen-related foods can cause late eczematous reactions in patients with atopic dermatitis. *Allergy* 2018;**73**:2046–54.
31. Ferreira MA, Vonk JM, Baurecht H, Marenholz I, Tian C, Hoffman JD, et al. Shared genetic origin of asthma, hay fever and eczema elucidates allergic disease biology. *Nat Genet* 2017;**49**:1752–7.
32. Wuthrich B, Schmid-Grendelmeier P. The atopic march. *Allergy* 2018;**73**:1753.
33. Khan SJ, Dharmage SC, Matheson MC, Gurrin LC. Response to “the atopic march”. *Allergy* 2018;**73**:1754.
34. Jatzlauk G, Bartel S, Heine H, Schlöter M, Krauss-Etschmann S. Influences of environmental bacteria and their metabolites on allergies, asthma, and host microbiota. *Allergy* 2017;**72**:1859–67.
35. Chu DM, Ma J, Prince AL, Antony KM, Seferovic MD, Aagaard KM. Maturation of the infant microbiome community structure and function across multiple body sites and in relation to mode of delivery. *Nat Med* 2017;**23**:314–26.
36. Torow N, Marsland BJ, Horne MW, Gollwitzer ES. Neonatal mucosal immunology. *Mucosal Immunol* 2017;**10**:5–17.
37. Fujimura KE, Sitarik AR, Havstad S, Lin DL, Levan S, Fadrosch D, et al. Neonatal gut microbiota associates with childhood multisensitized atopy and T cell differentiation. *Nat Med* 2016;**22**:1187–91.
38. Elbert NJ, van Meel ER, den Dekker HT, de Jong NW, Nijsten TEC, Jaddoe VWW, et al. Duration and exclusiveness of breastfeeding and risk of childhood atopic diseases. *Allergy* 2017;**72**:1936–43.
39. Gerlich J, Bencke N, Peters-Weist AS, Heinrich S, Roller D, Genuneit J, et al. Pregnancy and perinatal conditions and atopic disease prevalence in childhood and adulthood. *Allergy* 2018;**73**:1064–74.
40. Nicklaus S, Divaret-Chauveau A, Chardon ML, Roduit C, Kaulek V, Ksiazek E, et al. The protective effect of cheese consumption at 18 months on allergic diseases in the first 6 years. *Allergy* 2019;**74**:788–98.
41. Roduit C, Frei R, Ferstl R, Loeliger S, Westermann P, Rhyner C, et al. High levels of butyrate and propionate in early life are associated with protection against atopy. *Allergy* 2019;**74**:799–809.
42. Egeberg A, Andersen YM, Gislason GH, Skov L, Thyssen JP. Prevalence of comorbidity and associated risk factors in adults with atopic dermatitis. *Allergy* 2017;**72**:783–91.
43. Silverwood RJ, Forbes HJ, Abuabara K, Ascott A, Schmidt M, Schmidt SAJ, et al. Severe and predominantly active atopic eczema in adulthood and long term risk of cardiovascular disease: population based cohort study. *BMJ* 2018;**361**:k1786.
44. Lee S, Shin A. Association of atopic dermatitis with depressive symptoms and suicidal behaviors among adolescents in Korea: the 2013 Korean Youth Risk Behavior Survey. *BMC Psychiatry* 2017;**17**:3.
45. Lee SH, Lee SH, Lee SY, Lee B, Lee SH, Park YL. Psychological health status and health-related quality of life in adults with atopic dermatitis: a nationwide cross-sectional study in South Korea. *Acta Derm Venereol* 2018;**98**:89–97.
46. Thyssen JP, Hamann CR, Linneberg A, Dantoft TM, Skov L, Gislason GH, et al. Atopic dermatitis is associated with anxiety, depression, and suicidal ideation, but not with psychiatric hospitalization or suicide. *Allergy* 2018;**73**:214–20.
47. Sandhu JK, Wu KK, Bui TL, Armstrong AW. Association between atopic dermatitis and suicidality: a systematic review and meta-analysis. *JAMA Dermatol* 2019;**155**:178–87.
48. Ronnstad ATM, Halling-Overgaard AS, Hamann CR, Skov L, Egeberg A, Thyssen JP. Association of atopic dermatitis with depression, anxiety, and suicidal ideation in children and adults: a systematic review and meta-analysis. *J Am Acad Dermatol* 2018;**79**:448–56. e30.
49. Goldman LS, Genel M, Bezman RJ, Slanetz PJ. Diagnosis and treatment of attention-deficit/hyperactivity disorder in children and adolescents. Council on Scientific Affairs, American Medical Association. *JAMA* 1998;**279**:1100–7.
50. Sidbury R, Khorsand K. Evolving concepts in atopic dermatitis. *Curr Allergy Asthma Rep* 2017;**17**:42.
51. Miyazaki C, Koyama M, Ota E, Swa T, Mlunde LB, Amiya RM, et al. Allergic diseases in children with attention deficit hyperactivity disorder: a systematic review and meta-analysis. *BMC Psychiatry* 2017;**17**:120.
52. Johansson EK, Ballardini N, Kull I, Bergstrom A, Wahlgren CF. Association between preschool eczema and medication for attention-deficit/hyperactivity disorder in school age. *Pediatr Allergy Immunol* 2017;**28**:44–50.
53. Schmitt J, Buske-Kirschbaum A, Tesch F, Trikojatz K, Stephan V, Abraham S, et al. Increased attention-deficit/hyperactivity symptoms in atopic dermatitis are associated with history of antihistamine use. *Allergy* 2018;**73**:615–26.
54. Sokolowska M, Akdis CA. Highlights in immune response, microbiome and precision medicine in allergic disease and asthma. *Curr Opin Immunol* 2017;**48**:iv–ix.
55. Overgaard LEK, Main KM, Frederiksen H, Stender S, Szecsi PB, Williams HC, et al. Children with atopic dermatitis and frequent emollient use have increased urinary levels of low-molecular-weight phthalate metabolites and parabens. *Allergy* 2017;**72**:1768–77.
56. Goleva E, Berdyshev E, Leung DY. Epithelial barrier repair and prevention of allergy. *J Clin Invest* 2019;**129**:1463–74.
57. Akdis CA, Akdis M. Immunological differences between intrinsic and extrinsic types of atopic dermatitis. *Clin Exp Allergy* 2003;**33**:1618–21.
58. Rebane A, Akdis CA. MicroRNAs in allergy and asthma. *Curr Allergy Asthma Rep* 2014;**14**:424.
59. Karner J, Wawrzyniak M, Tankov S, Runnel T, Aints A, Kisand K, et al. Increased microRNA-323-3p in IL-22/IL-17-producing T cells and asthma: a role in the regulation of the TGF-beta pathway and IL-22 production. *Allergy* 2017;**72**:55–65.
60. Carreras-Badosa G, Runnel T, Plaas M, Karner J, Ruckert B, Lattekivi F, et al. microRNA-146a is linked to the production of IgE in mice but not in atopic dermatitis patients. *Allergy* 2018;**73**:2400–3.
61. Mitamura Y, Nunomura S, Nanri Y, Ogawa M, Yoshihara T, Masuoka M, et al. The IL-13/periostin/IL-24 pathway causes epidermal barrier dysfunction in allergic skin inflammation. *Allergy* 2018;**73**:1881–91.
62. Izuhara K, Nunomura S, Nanri Y, Ono J, Takai M, Kawaguchi A. Periostin: an emerging biomarker for allergic diseases. *Allergy* 2019. <https://doi.org/10.1111/all.13814>.
63. Fuhlbrigge RC, Kieffer JD, Armerding D, Kupper TS. Cutaneous lymphocyte antigen is a specialized form of PSGL-1 expressed on skin-homing T cells. *Nature* 1997;**389**:978–81.
64. Akdis M, Simon HU, Weigl L, Kreyden O, Blaser K, Akdis CA. Skin homing (cutaneous lymphocyte-associated antigen-positive) CD8+ T cells respond to superantigen and contribute to eosinophilia and IgE production in atopic dermatitis. *J Immunol* 1999;**163**:466–75.
65. Czarnecki T, Santamaria-Babi LF, Guttman-Yassky E. Circulating CLA(+) T cells in atopic dermatitis and their possible role as peripheral biomarkers. *Allergy* 2017;**72**:366–72.
66. Brunner PM, Emerson RO, Tipton C, Garcet S, Khattri S, Coats I, et al. Nonlesional atopic dermatitis skin shares similar T-cell clones with lesional tissues. *Allergy* 2017;**72**:2017–25.
67. Ito Y, Satoh T, Takayama K, Miyagishi C, Walls AF, Yokozeki H. Basophil recruitment and activation in inflammatory skin diseases. *Allergy* 2011;**66**:1107–13.
68. Gschwandtner M, Paulitschke V, Mildner M, Brunner PM, Hacker S, Eisenwort G, et al. Proteome analysis identifies L1CAM/CD171 and DPP4/CD26 as novel markers of human skin mast cells. *Allergy* 2017;**72**:85–97.
69. Wedman PA, Aladhami A, Chumanevich AP, Fuseler JW, Oskeritzian CA. Mast cells and sphingosine-1-phosphate underlie prelesional remodeling in a mouse model of eczema. *Allergy* 2018;**73**:405–15.
70. Kortekaas Krohn I, Shikhagaie MM, Golebski K, Bernink JH, Breynaert C, Creyns B, et al. Emerging roles of innate lymphoid cells in inflammatory diseases: clinical implications. *Allergy* 2018;**73**:837–50.
71. Furue M, Yamamura K, Kido-Nakahara M, Nakahara T, Fukui Y. Emerging role of interleukin-31 and interleukin-31 receptor in pruritus in atopic dermatitis. *Allergy* 2018;**73**:29–36.
72. Otsuka M, Egawa G, Kabashima K. Uncovering the mysteries of Langerhans cells, inflammatory dendritic epidermal cells, and monocyte-derived Langerhans cell-like cells in the epidermis. *Front Immunol* 2018;**9**:1768.
73. Wollenberg A, Kraft S, Hanau D, Bieber T. Immunomorphological and ultrastructural characterization of Langerhans cells and a novel, inflammatory dendritic epidermal cell (IDEC) population in lesional skin of atopic eczema. *J Invest Dermatol* 1996;**106**:446–53.
74. Iwamoto K, Numm TJ, Koch S, Herrmann N, Leib N, Bieber T. Langerhans and inflammatory dendritic epidermal cells in atopic dermatitis are tolerated toward TLR2 activation. *Allergy* 2018;**73**:2205–13.
75. Koch S, Stroisch TJ, Vorac J, Herrmann N, Leib N, Schnautz S, et al. AhR mediates an anti-inflammatory feedback mechanism in human Langerhans cells involving Fcεpsilon RI and Ido. *Allergy* 2017;**72**:1686–93.
76. Globinska A, Boonpiyathad T, Satitsuksanoa P, Kleuskens M, van de Veen W, Sokolowska M, et al. Mechanisms of allergen-specific immunotherapy: diverse mechanisms of immune tolerance to allergens. *Ann Allergy Asthma Immunol* 2018;**121**:306–12.
77. Baek JO, Lee JR, Roh JY, Jung Y. Oral tolerance modulates the skin transcriptome in mice with induced atopic dermatitis. *Allergy* 2018;**73**:962–6.
78. Baek JO, Roh JY, Jung Y. Oral tolerance inhibits atopic dermatitis-like type 2 inflammation in mice by modulating immune microenvironments. *Allergy* 2017;**72**:397–406.
79. Ventre E, Rozieres A, Lenief V, Albert F, Rossio P, Laoubi L, et al. Topical ivermectin improves allergic skin inflammation. *Allergy* 2017;**72**:1212–21.
80. Song JY, Kang HJ, Ju HM, Park A, Park H, Hong JS, et al. Umbilical cord-derived mesenchymal stem cell extracts ameliorate atopic dermatitis in mice by reducing the T cell responses. *Sci Rep* 2019;**9**:6623.
81. Sah SK, Agrahari G, Nguyen CT, Kim YS, Kang KS, Kim TY. Enhanced therapeutic effects of human mesenchymal stem cells transduced with superoxide dismutase 3 in a murine atopic dermatitis-like skin inflammation model. *Allergy* 2018;**73**:2364–76.
82. Rubic-Schneider T, Carballido-Perrig N, Regairaz C, Raad L, Jost S, Rauld C, et al. GPR91 deficiency exacerbates allergic contact dermatitis while reducing arthritic disease in mice. *Allergy* 2017;**72**:444–52.
83. Kopfnagel V, Wagenknecht S, Brand L, Zeitvogel J, Harder J, Hofmann K, et al. RNase 7 downregulates TH2 cytokine production by activated human T cells. *Allergy* 2017;**72**:1694–703.
84. Castan L, Magnan A, Bouchaud G. Chemokine receptors in allergic diseases. *Allergy* 2017;**72**:682–90.
85. Murray C, Ahrens K, Devalaraja M, Dymond M, Fagura M, Hargreaves A, et al. Use of a canine model of atopic dermatitis to investigate the efficacy of a CCR4 antagonist in allergen-induced skin inflammation in a randomized study. *J Invest Dermatol* 2016;**136**:665–71.

86. Xia M, Hu S, Fu Y, Jin W, Yi Q, Matsui Y, et al. CCR10 regulates balanced maintenance and function of resident regulatory and effector T cells to promote immune homeostasis in the skin. *J Allergy Clin Immunol* 2014;**134**: 634–44. e10.
87. Thostesen LM, Kjaergaard J, Pihl GT, Birk NM, Nissen TN, Aaby P, et al. Neonatal BCG vaccination and atopic dermatitis before 13 months of age: a randomized clinical trial. *Allergy* 2018;**73**:498–504.
88. Akdis CA, Ballas ZK. Precision medicine and precision health: building blocks to foster a revolutionary health care model. *J Allergy Clin Immunol* 2016;**137**: 1359–61.
89. Agache I, Akdis CA. Endotypes of allergic diseases and asthma: an important step in building blocks for the future of precision medicine. *Allergol Int* 2016;**65**:243–52.
90. Muraro A, Lemanske Jr RF, Hellings PW, Akdis CA, Bieber T, Casale TB, et al. Precision medicine in patients with allergic diseases: airway diseases and atopic dermatitis-PRACTALL document of the European Academy of allergy and clinical immunology and the American Academy of allergy, asthma & immunology. *J Allergy Clin Immunol* 2016;**137**:1347–58.
91. Muraro A, Fokkens WJ, Pietikainen S, Borrelli D, Agache I, Bousquet J, et al. European symposium on precision medicine in allergy and airways diseases: report of the European Union parliament symposium (October 14, 2015). *Allergy* 2016;**71**:583–7.
92. Muraro A, Lemanske Jr RF, Castells M, Torres MJ, Khan D, Simon HU, et al. Precision medicine in allergic disease-food allergy, drug allergy, and anaphylaxis-PRACTALL document of the European Academy of allergy and clinical immunology and the American Academy of allergy, asthma and immunology. *Allergy* 2017;**72**:1006–21.
93. Platts-Mills TAE, Schuyler AJ, Erwin EA, Commins SP, Woodfolk JA. IgE in the diagnosis and treatment of allergic disease. *J Allergy Clin Immunol* 2016;**137**: 1662–70.
94. Straumann A, Blanchard C, Radonjic-Hoesli S, Bussmann C, Hruz P, Saffronova E, et al. A new eosinophilic esophagitis (EoE)-like disease without tissue eosinophilia found in EoE families. *Allergy* 2016;**71**:889–900.
95. Mukai K, Gaudenzio N, Gupta S, Vivanco N, Bendall SC, Maecker HT, et al. Assessing basophil activation by using flow cytometry and mass cytometry in blood stored 24 hours before analysis. *J Allergy Clin Immunol* 2017;**139**: 889–99. e11.
96. Schussler E, Kattan J. Allergen component testing in the diagnosis of food allergy. *Curr Allergy Asthma Rep* 2015;**15**:55.
97. Flores Kim J, McCleary N, Nwaru BI, Stoddart A, Sheikh A. Diagnostic accuracy, risk assessment, and cost-effectiveness of component-resolved diagnostics for food allergy: a systematic review. *Allergy* 2018;**73**:1609–21.
98. Beyer K, Grabenhenrich L, Hartl M, Beder A, Kalb B, Ziegert M, et al. Predictive values of component-specific IgE for the outcome of peanut and hazelnut food challenges in children. *Allergy* 2015;**70**:90–8.
99. Lange L, Lasota L, Finger A, Vlainic D, Busing S, Meister J, et al. Ana o 3-specific IgE is a good predictor for clinically relevant cashew allergy in children. *Allergy* 2017;**72**:598–603.
100. Datema MR, van Ree R, Asero R, Barreales L, Belohlavkova S, de Blay F, et al. Component-resolved diagnosis and beyond: multivariable regression models to predict severity of hazelnut allergy. *Allergy* 2018;**73**:549–59.
101. Du Toit G, Roberts G, Sayre PH, Bahnson HT, Radulovic S, Santos AF, et al. Randomized trial of peanut consumption in infants at risk for peanut allergy. *N Engl J Med* 2015;**372**:803–13.
102. Hong X, Hao K, Ladd-Acosta C, Hansen KD, Tsai HJ, Liu X, et al. Genome-wide association study identifies peanut allergy-specific loci and evidence of epigenetic mediation in US children. *Nat Commun* 2015;**6**:6304.
103. Hong X, Ladd-Acosta C, Hao K, Sherwood B, Ji H, Keet CA, et al. Epigenome-wide association study links site-specific DNA methylation changes with cow's milk allergy. *J Allergy Clin Immunol* 2016;**138**:908–11. e9.
104. Berings M, Karaaslan C, Altunbulakli C, Gevaert P, Akdis M, Bachert C, et al. Advances and highlights in allergen immunotherapy: on the way to sustained clinical and immunologic tolerance. *J Allergy Clin Immunol* 2017;**140**: 1250–67.
105. Kollmann D, Nagl B, Ebner C, Emminger W, Wohrl S, Kitzmuller C, et al. The quantity and quality of alpha-gal-specific antibodies differ in individuals with and without delayed red meat allergy. *Allergy* 2017;**72**:266–73.
106. Matricardi PM, Kleine-Tebbe J, Hoffmann HJ, Valenta R, Hilger C, Hofmaier S, et al. EAACI molecular allergology user's guide. *Pediatr Allergy Immunol* 2016;**27**(Suppl 23):1–250.
107. Mothes-Luksch N, Raith M, Stingl G, Focke-Tejkl M, Razzazi-Fazeli E, Ziegelmayer R, et al. Pru p 3, a marker allergen for lipid transfer protein sensitization also in Central Europe. *Allergy* 2017;**72**:1415–8.
108. Wittenberg M, Nassiri M, Francuzik W, Lehmann K, Babina M, Worm M. Serum levels of 9alpha,11beta-PGF2 and apolipoprotein A1 achieve high predictive power as biomarkers of anaphylaxis. *Allergy* 2017;**72**:1801–5.
109. Pettersson ME, Koppelman GH, Flokstra-de Blok BMJ, Kollen BJ, Dubois AEJ. Prediction of the severity of allergic reactions to foods. *Allergy* 2018;**73**: 1532–40.
110. Blumchen K, Beder A, Beschornor J, Ahrens F, Gruebl A, Hamelmann E, et al. Modified oral food challenge used with sensitization biomarkers provides more real-life clinical thresholds for peanut allergy. *J Allergy Clin Immunol* 2014;**134**:390–8.
111. Baumert JL, Taylor SL, Koppelman SJ. Quantitative assessment of the safety benefits associated with increasing clinical peanut thresholds through immunotherapy. *J Allergy Clin Immunol Pract* 2018;**6**:457–65. e4.
112. Turner PJ, Wainstein BK. Crossing the threshold: can outcome data from food challenges be used to predict risk of anaphylaxis in the community? *Allergy* 2017;**72**:9–12.
113. Kleine-Tebbe J, Hamilton RG. Cashew allergy, 2S albumins, and risk predictions based on IgE antibody levels. *Allergy* 2017;**72**:515–8.
114. Alpan O, Loizou D, Santos I, Ness B, Plandowski J. Impact of immune work-up on outcomes and the cost of care in patients with Chronic Rhinosinusitis. *Allergy* 2019;**74**:1802–5.
115. Grabenhenrich LB, Reich A, Bellach J, Trendelenburg V, Sprickelmann AB, Roberts G, et al. A new framework for the documentation and interpretation of oral food challenges in population-based and clinical research. *Allergy* 2017;**72**:453–61.
116. Ronborg SM, Mosbech H, Poulsen LK. Exposure chamber for allergen challenge. A placebo-controlled, double-blind trial in house-dust-mite asthma. *Allergy* 1997;**52**:821–8.
117. Hashiguchi K, Tang H, Fujita T, Tsubaki S, Fujita M, Suematsu K, et al. Preliminary study on Japanese cedar pollinosis in an artificial exposure chamber (Ohio Chamber). *Allergol Int* 2007;**56**:125–30.
118. Hohlfield JM, Holland-Letz T, Larbig M, Lavae-Mokhtari M, Wierenga E, Kapsenberg M, et al. Diagnostic value of outcome measures following allergen exposure in an environmental challenge chamber compared with natural conditions. *Clin Exp Allergy* 2010;**40**:998–1006.
119. Lueer K, Biller H, Casper A, Windt H, Mueller M, Badorrek P, et al. Safety, efficacy and repeatability of a novel house dust mite allergen challenge technique in the Fraunhofer allergen challenge chamber. *Allergy* 2016;**71**: 1693–700.
120. Devillier P, Le Gall M, Horak F. The allergen challenge chamber: a valuable tool for optimizing the clinical development of pollen immunotherapy. *Allergy* 2011;**66**:163–9.
121. Corren J, Wood RA, Patel D, Zhu J, Yegin A, Dhilon G, et al. Effects of omalizumab on changes in pulmonary function induced by controlled cat room challenge. *J Allergy Clin Immunol* 2011;**127**:398–405.
122. Meyer W, Narkus A, Salapatek AM, Hafner D. Double-blind, placebo-controlled, dose-ranging study of new recombinant hypoallergenic Bet v 1 in an environmental exposure chamber. *Allergy* 2013;**68**:724–31.
123. Werfel T, Heratizadeh A, Niebuhr M, Kapp A, Roesner LM, Karch A, et al. Exacerbation of atopic dermatitis on grass pollen exposure in an environmental challenge chamber. *J Allergy Clin Immunol* 2015;**136**:96–103. e9.
124. Rosner-Friese K, Kaul S, Vieths S, Pfaar O. Environmental exposure chambers in allergen immunotherapy trials: current status and clinical validation needs. *J Allergy Clin Immunol* 2015;**135**:636–43.
125. Pfaar O, Demoly P, Gerth van Wijk R, Bonini S, Bousquet J, Canonica GW, et al. Recommendations for the standardization of clinical outcomes used in allergen immunotherapy trials for allergic rhinoconjunctivitis: an EAACI Position Paper. *Allergy* 2014;**69**:854–67.
126. Pfaar O, Calderon MA, Andrews CP, Angeli E, Bergmann KC, Bonlokke JH, et al. Allergen exposure chambers: harmonizing current concepts and projecting the needs for the future - an EAACI Position Paper. *Allergy* 2017;**72**:1035–42.
127. Jensen-Jarolim E, Pali-Scholl I, Roth-Walter F. Outstanding animal studies in allergy II. From atopic barrier and microbiome to allergen-specific immunotherapy. *Curr Opin Allergy Clin Immunol* 2017;**17**:180–7.
128. Novak N, Bieber T, Hoffmann M, Folster-Holst R, Homey B, Werfel T, et al. Efficacy and safety of subcutaneous allergen-specific immunotherapy with depigmented polymerized mite extract in atopic dermatitis. *J Allergy Clin Immunol* 2012;**130**:925–31. e4.
129. Bae JM, Choi YY, Park CO, Chung KY, Lee KH. Efficacy of allergen-specific immunotherapy for atopic dermatitis: a systematic review and meta-analysis of randomized controlled trials. *J Allergy Clin Immunol* 2013;**132**: 110–7.
130. Lee J, Park CO, Lee KH. Specific immunotherapy in atopic dermatitis. *Allergy Asthma Immunol Res* 2015;**7**:221–9.
131. Shin JU, Kim SH, Noh JY, Kim JH, Kim HR, Jeong KY, et al. Allergen-specific immunotherapy induces regulatory T cells in an atopic dermatitis mouse model. *Allergy* 2018;**73**:1801–11.
132. Rinaldi AO, Morita H, Wawrzyniak P, Dreher A, Grant S, Svedenhag P, et al. Direct assessment of skin epithelial barrier by electrical impedance spectroscopy. *Allergy* 2019. <https://doi.org/10.1111/all.13824>.
133. Bogh KL, van Bilsen J, Glogowski R, Lopez-Exposito I, Bouchaud G, Blanchard C, et al. Current challenges facing the assessment of the allergenic capacity of food allergens in animal models. *Clin Transl Allergy* 2016;**6**:21.
134. Jensen-Jarolim E, Pali-Scholl I, Roth-Walter F. Outstanding animal studies in allergy I. From asthma to food allergy and anaphylaxis. *Curr Opin Allergy Clin Immunol* 2017;**17**:169–79.
135. Jensen-Jarolim E, Einhorn L, Herrmann I, Thalhammer JG, Panakova L. Pollen allergies in humans and their dogs, cats and horses: differences and similarities. *Clin Transl Allergy* 2015;**5**:15.
136. Pali-Scholl I, De Lucia M, Jackson H, Janda J, Mueller RS, Jensen-Jarolim E. Comparing immediate-type food allergy in humans and companion animals-revealing unmet needs. *Allergy* 2017;**72**:1643–56.



137. Gaschen FP, Merchant SR. Adverse food reactions in dogs and cats. *Vet Clin North Am Small Anim Pract* 2011;**41**:361–79.
138. Tater KC, Jackson HA, Paps J, Hammerberg B. Effects of routine prophylactic vaccination or administration of aluminum adjuvant alone on allergen-specific serum IgE and IgG responses in allergic dogs. *Am J Vet Res* 2005;**66**:1572–7.
139. Oropeza AR, Bindslev-Jensen C, Broesby-Olsen S, Kristensen T, Moller MB, Vestergaard H, et al. Patterns of anaphylaxis after diagnostic workup: a follow-up study of 226 patients with suspected anaphylaxis. *Allergy* 2017;**72**:1944–52.
140. Kelleher MM, Dunn-Galvin A, Gray C, Murray DM, Kiely M, Kenny L, et al. Skin barrier impairment at birth predicts food allergy at 2 years of age. *J Allergy Clin Immunol* 2016;**137**:1111–6. e8.
141. Plunkett CH, Nagler CR. The influence of the microbiome on allergic sensitization to food. *J Immunol* 2017;**198**:581–9.
142. Trendelenburg V, Tschirner S, Niggemann B, Beyer K. Hen's egg allergen in house and bed dust is significantly increased after hen's egg consumption-A pilot study. *Allergy* 2018;**73**:261–4.
143. Oyoshi MK, Oettgen HC, Chatila TA, Geha RS, Bryce PJ. Food allergy: insights into etiology, prevention, and treatment provided by murine models. *J Allergy Clin Immunol* 2014;**133**:309–17.
144. Lexmond WS, Goettel JA, Lyons JJ, Jacobse J, Deken MM, Lawrence MG, et al. FOXP3+ Tregs require WASP to restrain Th2-mediated food allergy. *J Clin Invest* 2016;**126**:4030–44.
145. Liu ZQ, Li XX, Qiu SQ, Yu Y, Li MG, Yang LT, et al. Vitamin D contributes to mast cell stabilization. *Allergy* 2017;**72**:1184–92.
146. Molloy J, Koplin JJ, Allen KJ, Tang MLK, Collier F, Carlin JB, et al. Vitamin D insufficiency in the first 6 months of infancy and challenge-proven IgE-mediated food allergy at 1 year of age: a case-cohort study. *Allergy* 2017;**72**:1222–31.
147. Sampson HA, O'Mahony L, Burks AW, Plaut M, Lack G, Akdis CA. Mechanisms of food allergy. *J Allergy Clin Immunol* 2018;**141**:11–9.
148. van Ginkel CD, Pettersson ME, Dubois AEJ, Koppelman GH. Association of STAT6 gene variants with food allergy diagnosed by double-blind placebo-controlled food challenges. *Allergy* 2018;**73**:1337–41.
149. Savage JH, Lee-Sarwar KA, Sordillo J, Bunyavanich S, Zhou Y, O'Connor G, et al. A prospective microbiome-wide association study of food sensitization and food allergy in early childhood. *Allergy* 2018;**73**:145–52.
150. Asai Y, Eslami A, van Ginkel CD, Akhabir L, Wan M, Ellis G, et al. Genome-wide association study and meta-analysis in multiple populations identifies new loci for peanut allergy and establishes C11orf30/EMSY as a genetic risk factor for food allergy. *J Allergy Clin Immunol* 2018;**141**:991–1001.
151. Berni Canani R, Paparo L, Nocerino R, Cosenza L, Pezzella V, Di Costanzo M, et al. Differences in DNA methylation profile of Th1 and Th2 cytokine genes are associated with tolerance acquisition in children with IgE-mediated cow's milk allergy. *Clin Epigenetics* 2015;**7**:38.
152. Paparo L, Nocerino R, Cosenza L, Aitoro R, D'Argenio V, Del Monaco V, et al. Epigenetic features of FoxP3 in children with cow's milk allergy. *Clin Epigenetics* 2016;**8**:86.
153. D'Argenio V, Del Monaco V, Paparo L, De Palma FDE, Nocerino R, D'Alessio F, et al. Altered miR-193a-5p expression in children with cow's milk allergy. *Allergy* 2018;**73**:379–86.
154. Nwaru BI, Hickstein L, Panesar SS, Roberts G, Muraro A, Sheikh A, et al. Prevalence of common food allergies in Europe: a systematic review and meta-analysis. *Allergy* 2014;**69**:992–1007.
155. Wal JM. Bovine milk allergenicity. *Ann Allergy Asthma Immunol* 2004;**93**:S2–11.
156. van de Veen W, Akdis M. Role of IgG4 in IgE-mediated allergic responses. *J Allergy Clin Immunol* 2016;**138**:1434–5.
157. Caubet JC, Lin J, Ahrens B, Gimenez G, Bardina L, Niggemann B, et al. Natural tolerance development in cow's milk allergic children: IgE and IgG4 epitope binding. *Allergy* 2017;**72**:1677–85.
158. Hochwallner H, Schulmeister U, Swoboda I, Focke-Tejkl M, Reininger R, Civa V, et al. Infant milk formulas differ regarding their allergenic activity and induction of T-cell and cytokine responses. *Allergy* 2017;**72**:416–24.
159. Leonard SA, Caubet JC, Kim JS, Groetch M, Nowak-Węgrzyn A. Baked milk- and egg-containing diet in the management of milk and egg allergy. *J Allergy Clin Immunol Pract* 2015;**3**:13–23. quiz 4.
160. Remington BC, Westerhout J, Campbell DE, Turner PJ. Minimal impact of extensive heating of hen's egg and cow's milk in a food matrix on threshold dose-distribution curves. *Allergy* 2017;**72**:1816–9.
161. Guhl EE, Hofstetter G, Lengger N, Hemmer W, Ebner C, Froschl R, et al. IgE, IgG4 and IgA specific to Bet v 1-related food allergens do not predict oral allergy syndrome. *Allergy* 2015;**70**:59–66.
162. Vieths S, Scheurer S, Ballmer-Weber B. Current understanding of cross-reactivity of food allergens and pollen. *Ann N Y Acad Sci* 2002;**964**:47–68.
163. Ballmer-Weber BK, Wuthrich B, Wangorsch A, Fotisch K, Altmann F, Vieths S. Carrot allergy: double-blinded, placebo-controlled food challenge and identification of allergens. *J Allergy Clin Immunol* 2001;**108**:301–7.
164. Zulehner N, Nagl B, Briza P, Roulias A, Ballmer-Weber B, Zlabinger GJ, et al. Characterization of the T-cell response to Dau c 1, the Bet v 1-homolog in carrot. *Allergy* 2017;**72**:244–51.
165. Sampson HA, Aceves S, Bock SA, James J, Jones S, Lang D, et al. Food allergy: a practice parameter update-2014. *J Allergy Clin Immunol* 2014;**134**:1016–25. e43.
166. Burks AW, Sampson HA, Plaut M, Lack G, Akdis CA. Treatment for food allergy. *J Allergy Clin Immunol* 2018;**141**:1–9.
167. Pajno GB, Fernandez-Rivas M, Arasi S, Roberts G, Akdis CA, Alvaro-Lozano M, et al. EAACI Guidelines on allergen immunotherapy: IgE-mediated food allergy. *Allergy* 2018;**73**:799–815.
168. Rigbi NE, Goldberg MR, Levy MB, Nachshon L, Golobov K, Elizur A. Changes in patient quality of life during oral immunotherapy for food allergy. *Allergy* 2017;**72**:1883–90.
169. Polloni L, DunnGalvin A, Ferruzza E, Bonaguro R, Lazzarotto F, Toniolo A, et al. Coping strategies, alexithymia and anxiety in young patients with food allergy. *Allergy* 2017;**72**:1054–60.
170. Nurmatov U, Dhami S, Arasi S, Pajno GB, Fernandez-Rivas M, Muraro A, et al. Allergen immunotherapy for IgE-mediated food allergy: a systematic review and meta-analysis. *Allergy* 2017;**72**:1133–47.
171. Wood RA. Food allergen immunotherapy: current status and prospects for the future. *J Allergy Clin Immunol* 2016;**137**:973–82.
172. Wood RA, Kim JS, Lindblad R, Nadeau K, Henning AK, Dawson P, et al. A randomized, double-blind, placebo-controlled study of omalizumab combined with oral immunotherapy for the treatment of cow's milk allergy. *J Allergy Clin Immunol* 2016;**137**:1103–10. e11.
173. MacGinnitie AJ, Rachid R, Gragg H, Little SV, Lakin P, Cianferoni A, et al. Omalizumab facilitates rapid oral desensitization for peanut allergy. *J Allergy Clin Immunol* 2017;**139**:873–81. e8.
174. Ryan D, Gerth van Wijk R, Angier E, Kristiansen M, Zaman H, Sheikh A, et al. Challenges in the implementation of the EAACI AIT guidelines: a situational analysis of current provision of allergen immunotherapy. *Allergy* 2018;**73**:827–36.
175. Hoffmann HJ, Valovirta E, Pfaar O, Moingeon P, Schmid JM, Skaarup SH, et al. Novel approaches and perspectives in allergen immunotherapy. *Allergy* 2017;**72**:1022–34.
176. Pohlit H, Bellinghausen I, Frey H, Saloga J. Recent advances in the use of nanoparticles for allergen-specific immunotherapy. *Allergy* 2017;**72**:1461–74.
177. Pfaar O, Bonini S, Cardona V, Demoly P, Jakob T, Jutel M, et al. Perspectives in allergen immunotherapy: 2017 and beyond. *Allergy* 2018;**73**(Suppl 104):5–23.
178. Srivastava KD, Siefert A, Fahmy TM, Caplan MJ, Li XM, Sampson HA. Investigation of peanut oral immunotherapy with CpG/peanut nanoparticles in a murine model of peanut allergy. *J Allergy Clin Immunol* 2016;**138**:536–43. e4.
179. Smaldini PL, Trejo F, Cohen JL, Piaggio E, Docena GH. Systemic IL-2/anti-IL-2Ab complex combined with sublingual immunotherapy suppresses experimental food allergy in mice through induction of mucosal regulatory T cells. *Allergy* 2018;**73**:885–95.
180. Greenhawt M, Fleischer DM, Chan ES, Venter C, Stukus D, Gupta R, et al. LEAPing through the looking glass: secondary analysis of the effect of skin test size and age of introduction on peanut tolerance after early peanut introduction. *Allergy* 2017;**72**:1254–60.
181. Shaker M, Stukus D, Chan ES, Fleischer DM, Spergel JM, Greenhawt M. "To screen or not to screen": comparing the health and economic benefits of early peanut introduction strategies in five countries. *Allergy* 2018;**73**:1707–14.
182. Shaker M, Verma K, Greenhawt M. The health and economic outcomes of early egg introduction strategies. *Allergy* 2018;**73**:2214–23.